



Shannon pipeline plug-in: For human mRNA splicing mutations

CLC bio Genomics Workbench plug-in

CLC bio Genomics Server plug-in

Features and Benefits

Cytognomix introduces a line of Shannon pipeline plug-ins for prediction of functionally-significant, non-coding variants in genome or exome sequences. Comprehensive genome-scale analysis is now possible for mutations which completely or partially inactivate mRNA splice sites or activate cryptic splicing. The Shannon pipeline plug-in uses our patented and proven information theory-based binding site analysis. Its algorithm has been validated in hundreds of [peer-reviewed research studies of splicing mutations](#), and has been recognized by the American College of Medical Genetics and Genomics in their published guidelines and standards ([Genet. Med. 7, 571–583](#)).

The CLC Bio Genomics Workbench is used to visualize, export, and further analyze the results of the Shannon pipeline suite.

Cytognomix Tools

The CLC-Genomics Workbench retrieves lists of variants, and either processes the data itself or funnels it to the Shannon pipeline on the Genomics Server. Genome-wide information analysis is performed, then annotated against standard databases, and resulting mutations are filtered. Results are displayed as exportable Manhattan-style plots, sortable tables, or browser tracks.

User Benefits

- The Shannon Human Splicing Pipeline starts with hundreds of thousands of variants, then hones in on the very limited number that potentially alter mRNA splicing.
- Variants are categorized by whether they fully or partially inactivate natural sites or activate cryptic sites proximate to exons or within them.
- Predicts variants missed by many other techniques
- Splicing-related changes are displayed graphically either as “Manhattan-like” plots or as BED tracks.
- Mutations can be sorted according to:
 - the change in information content,
 - the proximity to a natural splice site,
 - the relative strength of cryptic vs adjacent natural site,
 - gene and chromosome coordinate.
- Variants that affect known single nucleotide polymorphisms (SNPs) are identified and can be filtered according to allele frequency.
- Results for genome-wide high throughput sequence data obtained in ≤ 2 hours.

- Fully integrated with CLC-Bio Genomics Workbench and Genomics Server

Features

- Identifies likely mutations with industry-leading sensitivity and specificity. Trusted legacy of experimentally validated mutation predictions.
- Input accepts variants in VCF, in *Cytognomix*'s simple indexed format, or as CLC Bio variant objects.
- Can accept variants and output results with either hg18 (NCBI36) or hg19 (GRCh37) coordinates.
- Intuitive exportable results based on sensible defaults.
- Run as a standalone application on the Genomics Workbench or configured as a client-server with both the Workbench and Server.
- Fully compatible output for other CLC Bio Workbench Tools.

Innovative platform for rapid, non-coding mutation analysis

Cytognomix uses information theory-based models of mRNA splicing to analyze mutations that alter transcript structure and abundance¹⁻³. Information models rank sequences according to their individual information content (R_i in bits). Functional binding sites have $R_i > 0$, corresponding to $\Delta G < 0$ kcal/mol. Strong binding sites have $R_i > R_{\text{sequence}}$ while weak sites have $R_i < R_{\text{sequence}}$. Variations which alter the affinity of a protein to bind there modify the R_i of the site. A 1 bit change in information content (ΔR_i) corresponds to a ≥ 2 fold change in binding affinity. This approach is applicable to any type of nucleic acid binding site, including transcription factors and other conserved non-coding sequences.

Predictions from these models are accurate²⁻⁴, as differences in individual information contents (ΔR_i in bits) are related to the splicesomal affinities of natural and variant sequences^{1,5}. Pathogenicity is related to ΔR_i , which is decreased at natural splice sites and/or increased at cryptic sites⁶⁻⁷. Sites with negative R_i values are not recognized. Leaky mutations have modestly reduced R_i values. Cryptic splice sites with R_i values exceeding adjacent natural splice sites are activated². The plug-in reports minimally detectable expression changes of >2 fold⁸, or $\Delta R_i > 1$ bit, as significant.

The Shannon pipeline was initially created to address the vexing problem of assessing the many variants of unknown significance that are detected in cancer genetic testing. The pipeline has been used to reanalyze the Breast Cancer Information Core identifying many splicing mutations, most of which were previously unrecognized⁹.

The Shannon software pipeline was developed and implemented in C and Perl to perform information analysis fast on a genome-wide scale. Determining R_i values of donor and acceptor sites along a nucleotide sequence is carried out using a convolution-style, sliding-window computation on chromosomes or subsets of chromosomes. R_i values are computed with $R_i(b,l)$ information weight matrix (based on a genome-wide set of verified donor and acceptor splice sites). Variants with significant ΔR_i values are then filtered, and annotated based on the gene they reside within. For

cryptic splice sites, the distance and relative location of the adjacent natural splice site of the same polarity is reported. Inactivating and leaky natural splice sites and cryptic splicing variants are categorized. Variants with known SNP designations and respective allele frequencies are also reported.

The CLC Bio implementation allows for additional filtering of input, and produces a graphical display of both ΔR_i and final R_i values for each variant on each chromosome, a table output which can be dynamically sorted that is categorized as separate tab for each type of mutation consequence, and BEDGRAPH output suitable for genome browsing. Results may be exported to a spreadsheet format for further data exploration.

References:

1. Schneider TD. J. Theor. Biol. 189: 427-41, 1997; 2. Rogan PK et al. Hum Mutat 12:153-171, 1998; 3. Rogan PK et al. Pharmacogenetics. 13:207-18, 2003; 4. Rogan PK, Schneider TD.. Hum Mutat 6:74-76, 1995; 5. Gadiraju S, et al. BMC Bioinformatics 4:38, 2003; 6. von Kodolitsch et al Circulation 100:693-9, 1999; 7. von Kodolitsch et al. 12: 258-262, 2006; 8. Nalla VK, Rogan PK. Hum Mut. 25(4):334-42, 2012.; 9. Mucaki et al. Hum. Mut. 32:735-742, 2012.

Benchmarks

The *Shannon Human Splicing Pipeline* analyzes an average of 3198 variants/min on an I7-based server:

Performance of Shannon pipeline plug-in on complete genome sequence data*

Number of variants	Complete analysis time
100,000	37 m
211,049	1h 12 m
290,589	1h 22 m
314,637	1h 27 m

*hg18/NCBI36

Requirements and validation

The Cytogenomix Shannon human mRNA splicing plug-in runs in standalone mode on the CLC Genomics Workbench V5.5 or with both the Workbench and CLC Genomics Server V.4.5 (as a standalone server or running Gridworks). Released for **Linux** and **MacOSX** Operating systems supporting Perl and gcc. Installation has been verified with Perl v.5.8.8 and 5.10.1 and gcc v.4.1.2 and v.4.4.3 with the Ubuntu 2.6.32-27 (32 and 64 bit), CentOS 2.6.18-238 (64 bit), and Fedora 16 (32 bit) kernels, and MacOSX (Lion release version 10.7.4; gcc v.4.2.1 and Perl 5.12.3) on hardware equipped with an Intel I7 processor and at least 4Gb RAM.

Support

CLC Bio customer support (primary)

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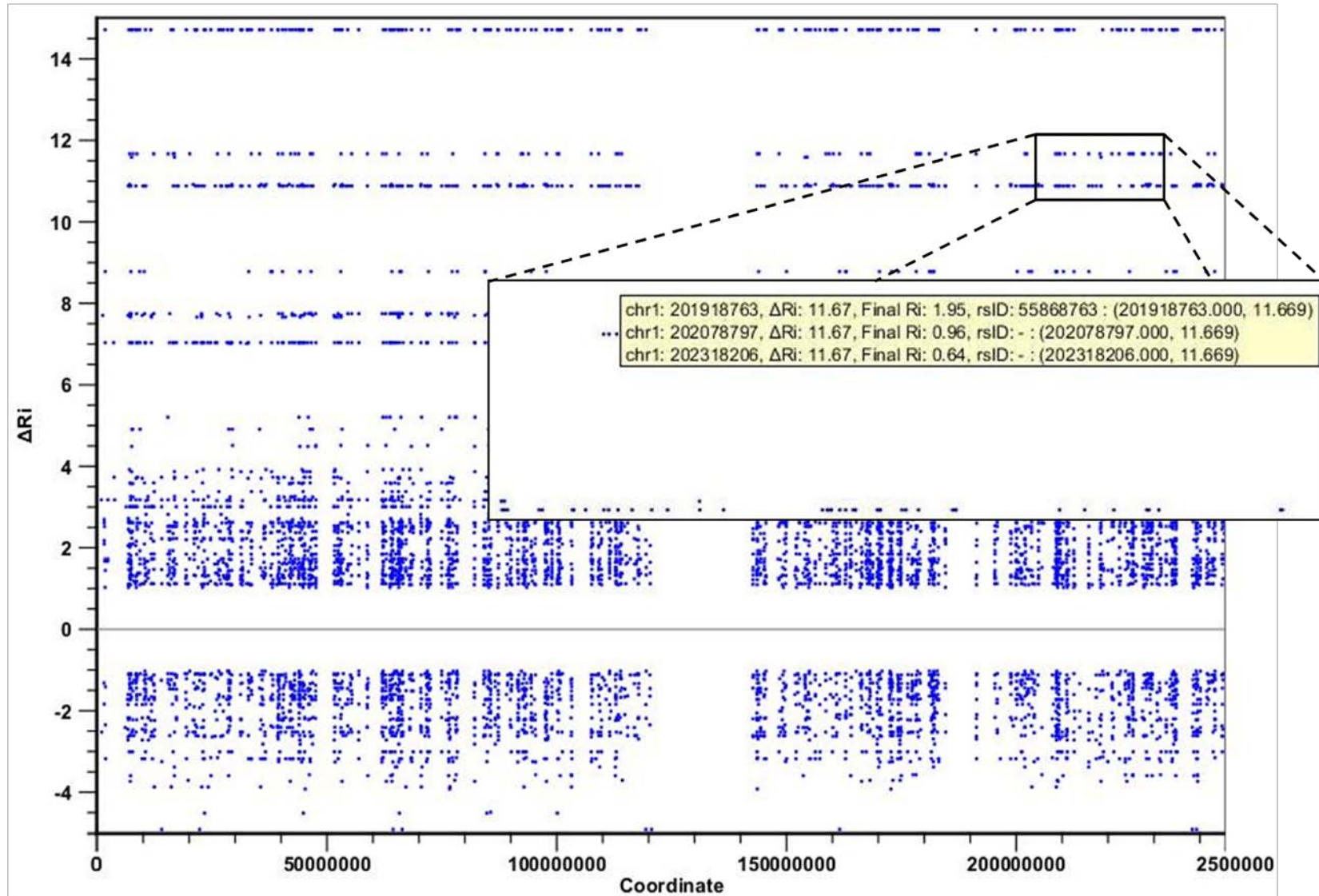
Tabular results from genome-wide mutation analysis produced by *CytognomiX's* Shannon mutation pipeline:

The screenshot displays the CLC Genomics Workbench 5.5 interface. The main window shows a table titled "Complete Vari..." with 7,073 rows. The table columns include: C..., Coordinate..., Ri-initial, Ri-final, ΔRi, Type, Gene Name, Location, Location..., Loc. Rel. ..., Dis..., Loc. of ne..., Ri of ..., and Cryptic Ri re. The table lists various genomic variants with their coordinates, Ri values, and associated gene names like C5orf17, HOTAIRM1, CACNA1B, ATRNL1, ARHGAP19, USH1C, OR51B6, RP11-65..., RP11-84..., UNC45A, NLR3, MYH4, AC08728..., AC08728..., SIGLEC1, SEC14L3, GSI-600..., F8, RP11-33..., RP11-14N..., RP11-26..., RP11-26..., RGS7, AC0994..., RIF1, DLEC1, CAMK1, PKX, AC09746..., HECW1, DUS4L, DAGLB, TUSC3, ZNF32-AS3, C10orf55, and YBRA1.

On the right side, the "Table Settings" panel is open, showing a list of columns to be displayed. The "Show column" section includes: Chromosome, Coordinate, Strand, Ri-initial, Ri-final, ΔRi, Type, Gene Name, Location, Location Type, Location Rel. to exon, Dist. from nearest nat. site, Loc. of nearest nat. site, Ri of nearest nat. site, Cryptic Ri relative to nat., rsID if available, Average heterozygosity, Input coordinate, Input variant, and Input ID. The "Column width" is set to "Manual".

At the bottom right of the interface, it indicates "0 rows selected".

``Manhattan``-like plot output of potential splicing mutations (unfiltered) on chromosome 1 generated by *CytognomiX*'s Shannon mutation pipeline. Inset shows context-dependent mutation detail generated by mouse-over:



Graphical custom genome browser track output produced by *Cytognomix's* Shannon mutation pipeline indicating information changes of a series of mutations occurring in the *BRCA1* gene (Mucaki et al. Hum. Mut. 32:735-742, 2012):

